PowerVision+TM Poly-AP IHC Detection System
(Biotin-free, anti-Mouse/Rabbit Primary Antibodies)

Cat No.	Alternative Cat. No.	For No. of Slides	Post-blocking (Polymer Penetration Enhancer)	Poly-AP anti-Mouse/Rabbit IgG
PV6102	DPVB+1000AP	10,000	1000 mL, RTU	1000 mL, RTU

I. INTENDED USE

For Research Use Only.
PowerVision+ Poly-AP IHC Detection Systems are intended for the chromogenic detection of targeted antigens that have been reacted to a user-supplied primary antibody. It is recommended that the reagents are not substituted across detection systems or lot numbers.

II. INTRODUCTION

PowerVision and PowerVision+ IHC Detection Systems utilize a novel poly-labeling technology, wherein secondary antibodies are directly polymerized with HRP or AP into compact polymers bearing higher ratio of enzymes to antibodies. These polymers demonstrated drastically improved detection sensitivity, efficiency and reliability comparing to conventional secondary antibody conjugates. Direct polymerization also avoids endogenous biotin reaction.

III. REAGENTS AND MATERIALS SUPPLIED

For exact catalog number of detection systems and their contents please refer to the above table.

Post-blocking (Polymer Penetration Enhancer)

Ready-to-use, a reagent that enhances Poly-AP anti-Mouse/Rabbit IgG polymer penetration and the interaction of polymers with primary antibodies.

Poly-AP anti-Mouse/Rabbit IgG

Ready-to-use, Poly-AP anti-Mouse/Rabbit IgG Polymer

IV. HANDLING, STORAGE AND SHELF LIFE

Storage Conditions: All reagents are to be stored at 2-8°C. Void after expiration date as specified on detection system/ reagent label.
Precautions: Specimens before and after fixation, and all materials exposed to them, should be handled as if capable of transmitting infection and disposed of with proper precautions. Some reagents in this detection system contain hazardous material. The user is advised to consult the MSDS for further information.

V. REAGENTS AND MATERIALS NEEDED BUT NOT SUPPLIED

- Universal IHC Blocking/Diluent
- Substrate/Chromogen
- Primary antibodies

VI. STAINING PROCEDURE

Each staining run should include both positive and negative tissue control slides to confirm
1. That the staining system is working properly
2. That positive and negative staining is specific
3. That the correct procedure has been followed.

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The combination of antigen retrieval protocol, primary antibody dilution, for use with a detection system should be determined by the user on a series of known positive and negative controls.

The tissue sections should not be allowed to dry out at any point during the staining procedures.
All procedures are performed at room temperature (18-26 °C).

1) Block with Universal IHC Blocking/Diluent (PV6123) for 10 min. Blot gently, no need to wash. Note: This step can be omitted if primary antibodies are diluted in the Universal IHC Blocking/Diluent
2) Apply primary antibodies for 30-60 min. Rinse well with buffer and wash in buffer for 5 min, twice
3) Apply Post-blocking and incubate for 20 min. Rinse well with buffer and soak in buffer for 5 min., twice.
4) Apply Poly-AP anti-Mouse/Rabbit IgG and incubate for 30 min. Rinse well with buffer and soak in buffer for 5 min., twice.
5) Apply Fast Red and incubate for 30 min. Rinse well with deionized or tap water
6) Counterstain and Mount: Proceed with appropriate counterstaining and mounting protocol.

VII. LIMITATIONS

Correct treatment of tissues prior to fixation and embedding is important for obtaining optimal results. Inconsistent results may be due to variation in fixation, embedding, pre-treatment and primary antibody reactivities, as well as from inherent variations in tissue. Leica Biosystems warrants that the materials sold meet our performance specifications until the expiration, if stored as recommended. No other warranties or guarantees, expressed or implied, are provided, including warranties for merchantability or fitness for a particular purpose.

VIII. GENERAL REFERENCE

1. S.R. Shi, J. Guo, R. Cote, L. Young, D. Hawes, Y. Shi, S. Thu and C. Taylor, “Sensitivity and Detection Efficiency of a Novel Biotin-free IHC Detection System: PowerVision”, Applied Immunohistochemistry & Molecular Morphology., 7:201-208, 1999
2. K. Petrosyan, R. Tamayo, D. Joseph, “Sensitivity of a Novel Biotin-free Detection Reagent (PowerVision+) for IHC” J. Histotechnology, 25:247-250, 2002
3. G. Bricca, et al., “Immunostaining Melanoma Frozen Sections: The 1-Hour Protocol” Dermatologic. Surgery, 30:403- 408, 2004

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